

## DESCRIPTION

Harris Hematoxylin is used for histology specimen staining; and this regressive stain appears darkest/most intense in appearance. Use this stain if the Gill 3X Hematoxylin staining is not providing a dark enough nuclear stain. Alternatively, use the Harris, Acidified as a progressive method for fewer steps and the same good quality.

## PROTOCOL FOR HARRIS H&E STAIN FOR HISTOLOGY

- Paraffin slides:** Deparaffinize in xylene, then rehydrate through graded alcohols to water.  
**Frozen slides:** Fix in 95% Ethanol (or other fixative according to lab preference) and rinse in water.
- Stain in Hematoxylin Solution, Harris: 4 to 6 minutes
- Rinse in two changes of DI water: 1 minute each  
*Note: this could also be done using running tap water, but we recommend DI water to avoid the varying pH found in tap water.*
- Acid Alcohol 0.5%, or Acid Alcohol 1%: 10 seconds  
*Note: Some prefer to use a weaker acid alcohol rinse which is 0.5% while others prefer the 1% acid alcohol rinse. The higher percent solution will extract dye a little fast but there is not much difference from the weaker solution here.*
- Rinse in two changes of DI water: 1 minute each
- Bluing Reagent (until tissue turns blue): 30 to 60 seconds
- Rinse in two changes of DI water: 1 minute each
- Ethanol 95%: 15 to 30 seconds
- Eosin Y 1% Alcoholic, or Eosin Y Intensified, or Eosin-Phloxine: 1 to 2 minutes  
*Note: For an increased definition of cytoplasmic components over Eosin Y 1%, select Eosin Y Intensified or Eosin-Phloxine as an alternative.*
- Dehydrate in Ethanol 95%, two changes: 10 dips each
- Dehydrate in Ethanol 100%, two changes: 1 minute each
- Clear in Xylene or Xylene substitute, two changes: 1 minute each
- Coverslip using Acrylic Mounting Medium and examined under microscope

## PROTOCOL FOR HARRIS HEMATOXYLIN, ACIDIFIED & EOSIN STAIN FOR HISTOLOGY

- Paraffin slides:** Deparaffinize in xylene, then rehydrate through graded alcohol to water.  
**Frozen slides:** Fix in Ethanol 95% (or other fixative according to lab preference) and rinse in water.
- Stain in Hematoxylin solution, Harris Hematoxylin, acidified 2 to 4 minutes
- Rinse in two changes of DI water: 1 minute each  
*Note: This could also be done using running tap water, but we recommend DI water to avoid the varying pH found in tap water.*
- Bluing Reagent (until tissue turns blue): 15 to 60 seconds
- Rinse in two changes of DI water: 30-60 seconds  
*Note: This could also be done using running tap water.*
- Ethanol 95%: 15 to 30 seconds
- Eosin Y 1% Alcoholic, Eosin Y Intensified, or Eosin-Phloxine: 30 to 60 seconds  
*Note: for an increased definition of cytoplasmic components over Eosin Y 1%, select Eosin Y Intensified or Eosin-Phloxine as a substitute.*
- Dehydrate in Ethanol 95%, two changes: 10 dips each
- Dehydrate in Ethanol 100%, two changes: 1 minute each
- Clear in Xylene or Xylene Substitute, two changes: 1 minute each.
- Coverslip using Acrylic Mounting Medium and examine under microscope

## RESULTS

Nuclei, nuclear components – blue to dark purple  
Cytoplasm, red blood cells, collagen, elastic fiber, muscle – shades of pink

## ORDERING INFORMATION

Cat. #	Description	Sizes
3315	Deionized Water	1 Gallon, 5 Gallon, 30 Gallon
7000	Hematoxylin Harris, Hg Free	16oz, 32oz, 1 Gallon
7002	Hematoxylin Harris, Hg Free, Acidified	16oz, 32oz, 1 Gallon, 5 Gallon
7560	Acid Alcohol, 0.5%, Histology	1 Gallon
7561	Acid Alcohol, 1%, Histology	1 Gallon
3356	Bluing Reagent	32oz, 1 Gallon
7006	Eosin Y, 1% Alcoholic	16oz, 32oz, 1 Gallon
7007	Eosin Y, Intensified	16oz, 32oz, 1 Gallon
7009	Eosin - Phloxine	16oz, 32oz, 1 Gallon, 2.5 Gallon
3340	Ethanol Solution, 95%	1 Gallon, 5 Gallon
3341	Ethanol Solution, 100%	1 Gallon, 5 Gallon
3346	Xylene	1 Gallon, 5 Gallon
3349	Acrylic Mounting Medium	4oz, 16oz

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