

DESCRIPTION

A simple procedure for differentiation of morphological cell types in peripheral blood, bone marrow and blood parasites. Wright Giemsa Stain clearly defines individual cells, their nuclear detail and the cytoplasmic structure. The staining characteristics are consistent to a traditional Romanowsky stain. Wright Giemsa Stain offers the flexibility of varying the staining intensity from the classical results to more basophilic staining result by varying the staining times.

STORAGE CONDITIONS

Store at controlled room temperature (15-30°C). Keep bottle tightly closed when not in use. Do not use after expiration date specified on bottle label. **Indication of deterioration:** Excessive precipitation in reagent is an indication of degradation.

SPECIMEN COLLECTION AND PREPARATION

Smears prepared from fresh uncoagulated venous, arterial blood, or bone marrow should be air-dried. If the smears are not prepared immediately, 2mgs of EDTA/mL of blood/bone marrow sample should be added to prevent coagulation.

PROCEDURE

Materials Provided:	Materials Not Provided:
Wright Giemsa Stain	Staining Rack
Phosphate Buffer pH 6.8	Microscope Slides
Rinse Solution pH 7.0	Immersion Oil

TECHNIQUE (Blood)

1. Set-up a staining rack over a sink.
2. Prepare slides as instructed under Specimen Collection.
3. Flood each specimen slide with ~1 mL or an adequate volume of the stain solution for a minimum of 2 minutes.
4. Repeat this procedure by adding an equal amount of Buffer directly to the stain solution on the slide.
5. Rock the slide gently for 30 to 60 seconds or by applying a current of air and allow to-stand for 2 additional minutes. Rinse the slides with Rinse Solution pH 7.0, wipe edges of excess stain and allow to air dry. Examine using immersion oil.

TECHNIQUE (Bone Marrow)

1. Set-up a staining rack over a sink.
2. Prepare slides as instructed under Specimen Collection.
3. Fix smears for 3 minutes in methanol
4. Flood slide for 4 minutes with the supplied Wright-Giemsa stain
5. Add an equal amount of the supplied pH 6.8 buffer to the slide for 2 minutes. Mix by rocking the slide. Drain slide.
6. Dilute WG stain with equal volume of buffer and transfer to the slide for 8 minutes.
7. Rinse slide for 30 seconds in Rinse Solution pH 7.0.

NOTE: Staining intensity may be varied by increasing the time in the Wright Giemsa Stain and phosphate buffer. For a more basophilic result (blue intensity), increase the time in the stain from approximately 2 minutes to 3 - 4 minutes and the buffer from 3 minutes to 5 minutes. Any variation of the procedure other than described above will effect the results.

Specimen	Nucleus	Granules	Cytoplasm
Erythrocytes	NA	NA	pink
Leucocytes Granular Polymorphonuclear			
Neutrophils	purple	red-lilac	light pink
Eosinophils	dark blue	red/orange	medium blue
Basophils	dark blue	dark purple	light blue
Leucocytes Nongranular			
Monocytes	violet	NA	light blue
Lymphocytes	violet	NA	medium blue
Platelets	NA	purple	lilac
Microorganisms	blue/violet	NA	NA
Spermatozoa	dark blue		

PRECAUTIONS

Wright Stain Solution contains methyl alcohol and is a Flammable Liquid.

DANGER – Poison May Be Fatal or Cause Blindness if Swallowed. Vapor is Harmful and Will Cause Eye Irritation. Do not breathe vapor. Avoid contact with eyes, skin, or clothing. Keep container closed. Use with adequate ventilation. Keep from heat, sparks and open flame. Wash thoroughly after handling. CANNOT BE MADE NONPOISONOUS.

First Aid: If swallowed, call a physician immediately. In case of contact with eyes, immediately flush with water for 15 minutes and notify a physician. Wash exposed areas thoroughly after handling.

All products are warranted to perform as described in their labeling and corresponding literature. All other warranties expressed or implied, including the warranty of merchantability and fitness for use are excluded. In no event shall Astral Diagnostics, Inc. be liable for any indirect or consequential damages.

ORDERING INFORMATION

Cat. #	Description	Sizes
5585	Wright Giemsa Stain Set	3 x 8oz

Order today at AstralDiagnostics.com